# **INSTRUCTIONS**



# **ProFoldin** Large-scale Preparative Protein Folding Column Sets

# **CATALOG NUMBERS**

<b>PFC01</b> (column # 1)	<b>PFC02</b> (column # 2)	PFC03
<b>PFC04</b> (column # 4)	<b>PFC05</b> (column # 5)	PFC06
<b>PFC07</b> (column # 7)	<b>PFC08</b> (column # 8)	PFC09
<b>PFC10</b> (column # 10)		

**PFC03** (column # 3) **PFC06** (column # 6) **PFC09** (column # 9)

## **INTRODUCTION**

The Large-scale Preparative Protein Folding Column Sets are used for preparative protein folding after the folding condition has been identified by the Spin-column Protein Folding Screen Kit (Catalog number SFC01-10). The column number represents the specific folding condition. Each Large-scale Preparative Protein folding Column Set includes 4 identical preparative protein folding columns and reagents for folding 10 to 20 mg of guanidine hydrochloride or urea-solubilized inclusion body proteins.

The Large-scale Preparative Protein Folding Column Sets (Catalog No. PFC01 to PFC10) includes 4 columns, 5.4 ml of Solution A, 15.4 ml of Solution C and 8.4 ml Solution B.

### PROTEIN FOLDING PROCEDURE

### Inclusion body preparation and solubilization

- 1. Resuspend the cell pellet in 20 ml of cell lysis buffer (20 mM Tris-HCl, pH 8, 100 mM NaCl, 2 mM DTT, 2 mM EDTA) for each liter of culture.
- 2. Break the cells by passing the cell suspension French Press twice and centrifuge the broken cell suspension at 20,000 rpm for 20 min.
- 3. Resuspend the pellet in the cell lysis buffer plus 1 % Triton-100 by stirring at 4°C for 1 to 2 hours and centrifuge the suspension at 20,000 rpm for 20 min. Discard the supernatant.
- 4. Wash the pellet in the cell lysis buffer without Triton by suspension and centrifugation.
- 5. Solubilize the inclusion bodies by stirring the pellet in 20 mM Tris-HCl, pH 8.0, 6 M guanidine hydrochloride (or 8 M urea), 10 mM DTT at room temperature for 2 hours. Then centrifuge the solubilization material at 30,000 rpm for 45 min. Save the supernatant as the solubilized inclusion bodies.

#### Protein folding using the large-scale preparative protein folding columns

The columns and reagents are cooled to 2 - 8°C. The experiment is performed in a cold room. An easy way to set up the columns is to use a test tube rack to hold the columns and place a tip box cover under the rack to receive the solution from the columns.

# **INSTRUCTIONS**



#### 1. Sample preparation

To make the loading sample, mix 5 ml of the solubilized inclusion bodies with 5 ml of Solution A. Incubate the mixture (the loading sample) for 5 min.

### 2. Column preparation

Cut off the column bottom pointing tips and let the buffer run through the columns.

### 3. Protein folding

- (1) load 2.5 ml of the loading sample per column. Let the sample completely run into the columns.
- (2) Elute the protein with 3.5 ml of Solution C per column. The solution C number matches the column number. Collect and incubate the eluent at 4°C for 2 to 4 hr. Discard the columns.
- (3) Mix 8 ml of Solution B<sup>(a)</sup> with the 14 ml of eluent from the 4 columns and incubate the solution at 4°C for 2 hr to overnight. Remove the precipitate (if there is any) by centrifugation. Note: If solution B forms precipitate during storage, warm it to room temperature to solubilize the precipitate, then cool it back to 4°C before use.

## PROTEIN PURIFICATION AFTER FOLDING

The folded protein can be purified by affinity, ion-exchange or gel filtration column chromatography. Some proteins are sensitive to low salt. To be cautious, brief dialysis of the protein solution against a buffer with a moderate salt concentration at 4°C is recommended. Following is a Q-Sepharose column purification protocol:

- Dialyze the protein solution against 40 volumes of 20 mM Tris-HCl, pH 8.5, 50 mM NaCl, 2 mM DTT, 2 mM EDTA buffer at 4°C for 2 to 4 hr. Remove any precipitates by centrifugation.
- (2) Equilibrate a Q-Sepharose column with a low salt buffer (the same as the dialysis buffer). Load the dialyzed protein solution. Wash the loaded column with 10 column volumes of the same buffer. Elute the protein with a salt gradient.
- (3) Any further protein purification step may follow as purification of regular soluble native proteins.

# **RELATED PRODUCTS**

Spin-column Protein Folding Screen Kit 96-well protein folding plate Spin-column Membrane Protein Folding Screen Kit Catalog number: SFC01-10 Catalog number: PFS096 Catalog number: MFC01-20