Bacillus subtilis
P*grac*100 Expression Vectors

Mo Bi Tec
MOLECULAR BIOTECHNOLOGY

Contents

1. Features

1.1. Recombinant protein production with *Bacillus subtilis*

- Non-pathogenic and considered as a GRAS organism (generally regarded as safe)
- *B. subtilis* has no significant bias in codon usage
- It can secrete proteins directly into the culture medium
- MoBiTec host strain for secretory protein production available: *B. subtilis* WB800N, an eightfold extracellular protease deficient strain.
- MoBiTec host strains for intracellular protein production available

1.2. P*grac*100 inducible expression vectors

- P*grac*100 vectors own the improved strong P*grac*100 promotor
- Expression of the gene of interest is controlled by the improved IPTG inducible P*grac*100 promotor and the corresponding repressor encoded by lacI
- This allows highly efficient extra- and intracellular production of recombinant proteins
- Vectors are structurally and segregationally stable
- Convenient cloning due to *B. subtilis* / *E. coli* shuttle vectors
- MoBiTec P*grac*100 vectors with 8xHis- and Strep-tag available

2. Introduction

Gram-positive bacteria are well known for their contributions to agricultural, medical and food biotechnology and for the production of recombinant proteins. Among them, *Bacillus subtilis* has been developed as an attractive host. At present, about 60% of the commercially available enzymes are produced by *Bacillus* species. *B. subtilis* is advantageous in that it is non-pathogenic, does not have significant bias in codon usage, and is capable of secreting functional proteins directly into the culture medium in large scale. A large body of information is available concerning genome sequence, transcription, translation, protein folding and secretion mechanisms, genetic manipulation, and large-scale fermentation.

With the *B. subtilis* P*grac*100 expression vectors MoBiTec offers an easy to handle tool for *B. subtilis* for high yield intracellular protein production and also for secretion of heterologous proteins into the culture medium in very high amount.

3. The P*grac*100 Expression Vectors

The P*grac*100 expression vectors are improved derivatives of the P*grac*01 expression vectors. They are *B. subtilis* / *E. coli* shuttle vectors, constructed for cloning in *E. coli* and high level expression of heterologous proteins with *B. subtilis*. The vectors are structurally and segregationally stable, using the theta-mode of replication within *B. subtilis* (Jannière et al., 1990; Titok et al., 2003).
All P*grac*100 vectors use the strong promoter preceding the *groESL* operon of *Bacillus subtilis* with improved regulatory elements fused to the *lac* operator allowing their induction by IPTG. Nucleotides were optimized at the conserved regions of the *groESL* promoter including the UP element, the −35 and the −15 region. Combination of these changes into one promoter enhanced the amount of recombinant proteins accumulating intracellularly up to about 30 % of the total cellular protein of *B. subtilis* (Phan *et al.*, 2011). In addition, the target proteins could be also expressed efficiently in *E. coli* in some cases.

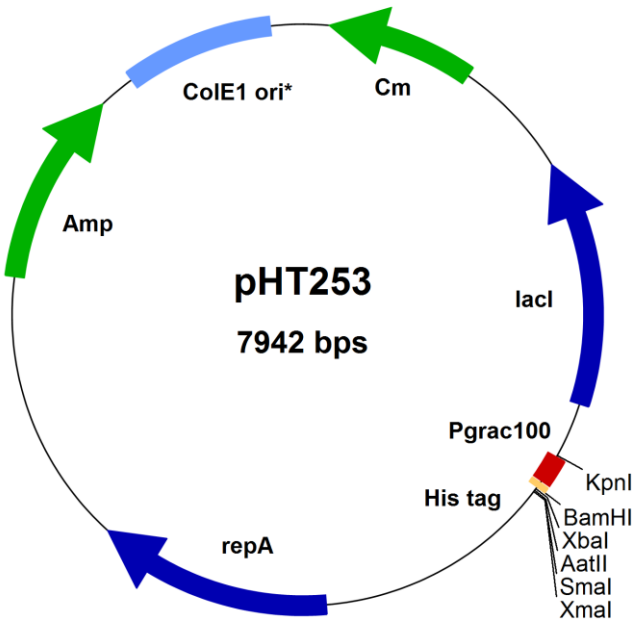
Vectors for intracellular protein production:

- pHT253: *Pgrac*100 expression vector with N-terminal encoded 8xHis tag
- pHT254: P*grac*100 expression vector with C-terminal encoded 8xHis tag
- pHT255: P*grac*100 expression vector with C-terminal encoded Strep tag

Vectors for secretory protein production:

- pHT1469: P*grac*100 expression vector with improved signal sequence of *amyQ* for protein export

3.1. Vector Map of pHT253

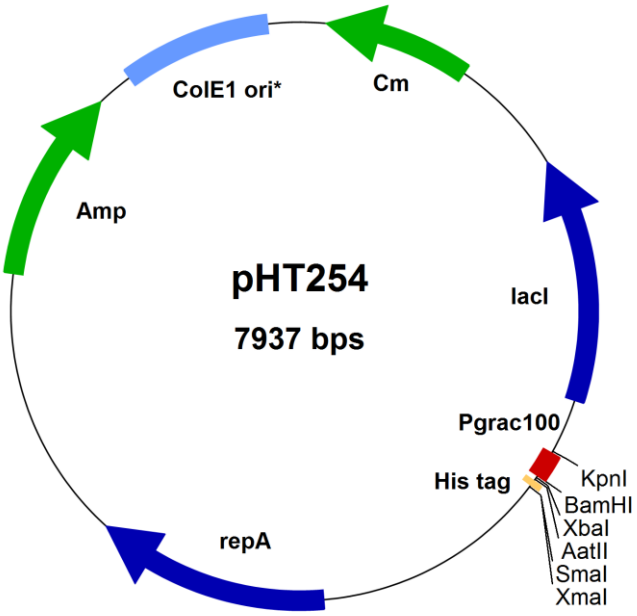


	Type	Start	End	Name	Description
	Selectable Genetic Marker	763	113	Cm	Chloramphenicol resistance (*B. subtilis*)
	Gene	2379	1297	lacI	lacI repressor gene
	Promoter	2631	2753	Pgrac100	P*grac100* promoter
	Tag	2763	2786	His tag	His tag
	Terminator	2819	2844	(not shown)	Region for transcription termination
	Gene	3869	4903	repA	replication gene A
	Selectable Genetic Marker	6124	6984	Amp	Ampicillin resistance gene (*E. coli*)
	Origin of replication	7146	7800	ColE1*	Origin of replication (*E. coli*); ColE1 incompatibility group

The sequence of the ribosome binding site (RBS), the multiple cloning site (MCS) and the position of the His tag in pHT253 is shown below. The complete DNA sequence is available at our website.

AAAGGAGGAAGGATCTATGGAAGCT-8xHis-tag- GGATCCATGTCTAGAGTCGACGTCCCCGGGGCAGCC
RBS BamHI XbaI AatII SmaI

3.2. Vector Map of pHT254

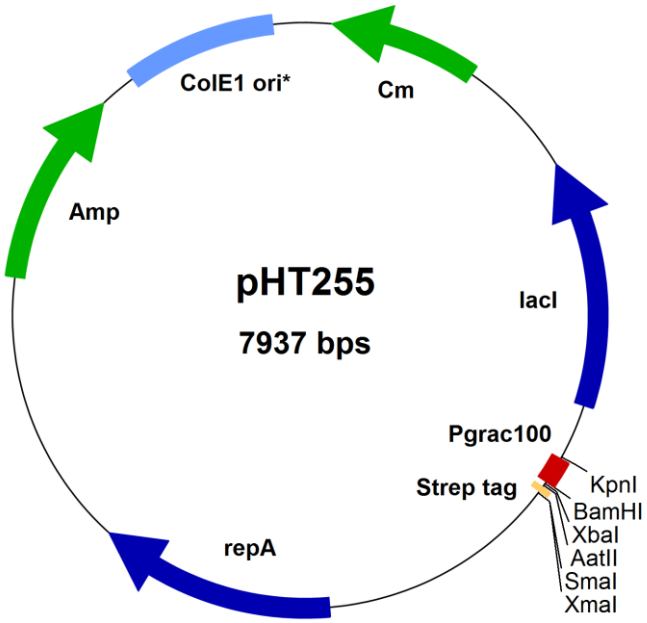


	Type	Start	End	Name	Description
	Selectable Genetic Marker	763	113	Cm	Chloramphenicol resistance (*B. subtilis*)
	Gene	2379	1297	lacI	lacI repressor gene
	Promoter	2631	2753	Pgrac100	P*grac100* promoter
	Tag	2774	2798	His tag	His tag
	Terminator	2814	2839	(not shown)	Region for transcription termination
	Gene	3864	4898	repA	replication gene A
	Selectable Genetic Marker	6119	6979	Amp	Ampicillin resistance gene (*E. coli*)
	Origin of replication	7141	7795	ColE1*	Origin of replication (*E. coli*); ColE1 incompatibility group

The sequence of the ribosome binding site (RBS), the multiple cloning site (MCS) and the position of the His tag in pHT254 is shown below. The complete DNA sequence is available at our website.

AAAGGAGGAAGGATCCATGTCTAGAGTCGACGTCGCT-8xHis-tag-TAACGTCCCCGGGGCAGCC
RBS BamHI XbaI AatII SmaI

3.3. Vector Map of pHT255

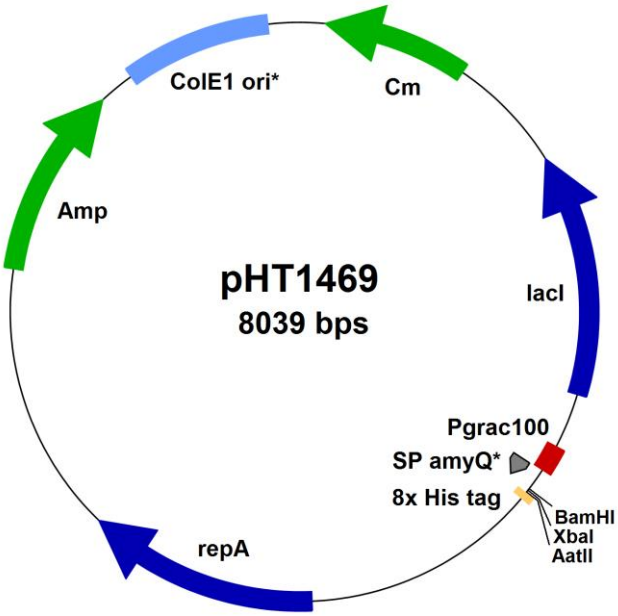


	Type	Start	End	Name	Description
	Selectable Genetic Marker	763	113	Cm	Chloramphenicol resistance (*B. subtilis*)
	Gene	2379	1297	lacI	lacI repressor gene
	Promoter	2631	2753	Pgrac100	P*grac100* promoter
	Tag	2772	2798	Strep tag	Strep tag
	Terminator	2814	2839	(not shown)	Region for transcription termination
	Gene	3864	4898	repA	replication gene A
	Selectable Genetic Marker	6119	6979	Amp	Ampicillin resistance gene (*E. coli*)
	Origin of replication	7141	7795	ColE1*	Origin of replication (*E. coli*); ColE1 incompatibility group

The sequence of the ribosome binding site (RBS), the multiple cloning site (MCS) and the position of the Strep tag in pHT255 is shown below. The complete DNA sequence is available at our website.

AAAGGAGGAA**GGATCC**A**TG**T**CTAGA**GTC**GACGTC**GCT-Strep-tag-**TAA**CGTC**CCCGGG**GCAGCC
RBS **BamHI** **XbaI** **AatII** **SmaI**

3.4. Vector Map of pHT1469



	Type	Start	End	Name	Description
	Selectable Genetic Marker	763	113	Cm	Chloramphenicol resistance (*B. subtilis*)
	Gene	2379	1297	lacI	lacI repressor gene
	Promoter	2631	2753	Pgrac100	P*grac100* promoter
	Signal Peptide	2754	2846	SP amyQ*	modified signal peptide of α-amylase
	Tag	2877	2903	His tag	His tag
	Terminator	2916	2941	(not shown)	Region for transcription termination
	Gene	3966	5000	repA	replication gene A
	Selectable Genetic Marker	6221	7081	Amp	Ampicillin resistance gene (*E. coli*)
	Origin of replication	7243	7897	ColE1*	Origin of replication (*E. coli*); ColE1 incompatibility group

The sequence of the ribosome binding site (RBS), the improved amyQ signal peptide (SP_{amyQ*}), and the multiple cloning site is shown below. The complete DNA sequence of pHT1469 is available at our website.

AAAGGAGGAAGGATCA ATG ATT CAA AAA CGA AAG CGG ACA GTT TCG TTC AGA CTT GTG CTT ATG
RBS M I Q K R K R T V S F R L V L M

TGC ACG CTG TTA TTT GTC AGT TTG CCG ATT ACA AAA GCA TCA GCT GCTGGATCCATGTCTAGAGTC
C T L L F V S L P I T K A S A BamHI XbaI

GACGTCGCT 8xHis tag-TAACGTCCCCGGGGCAGCC
AatII SmaI

4. *Bacillus subtilis* Host Strains

The following *Bacillus subtilis* strains suitable as hosts for gene expression are available:

For intracellular protein production:

- AS1: 1012 *hrcA*::*neo* (producing strain for enhancing solubility of intracellular protein Schulz and Schumann, 1996, and Phan *et al.*, 2006)
- 1012 wild type: *leuA8 metB5 trpC2 hsdRM1*
- 168 Marburg: *trpC2* (Trp^-)

For secretion vectors:

- WB800N: *nprE aprE epr bpr mpr::ble nprB::bsr Δvpr wprA::hyg cm::neo;* NeoR
  Please note that WB800N carries resistance to neomycin!

5. Storage and Handling Instructions

Storage and handling of plasmids

Plasmids are supplied lyophilized. Upon receipt, add 100 µl distilled water (final concentration 0.1 µg/µl) and incubate at 50 °C for 5 minutes. Vortex for 1 minute and store at -20 °C.

All plasmids of this system are *E. coli* / *B. subtilis* shuttle vectors.

Storage and handling of *Bacillus* strains

The *Bacillus* strains are supplied as frozen cultures and shipped on dry ice. Store the stock at -80 °C. For propagation remove tube from freezer, scratch off some material from the surface of the frozen stock using a sterile loop. Streak onto an LB plate, seal the plate with Parafilm and incubate at 37 °C overnight. *Bacillus* plates can be stored at 4 °C for 1 month. Use fresh bacteria for transformation.

6. Growth Conditions

Detailed protocols for *E. coli* and *Bacillus* molecular genetic handling (growth, transformation, etc.) can be found in the relevant laboratory manuals such as Sambrook and Russell (2001).
B. subtilis and *E. coli* can be grown aerobically at 37 °C in 2xYT medium (Bagyan *et al.*, 1998). Under optimal conditions the doubling time of *E. coli* is 20 min, of *B. subtilis* 30 min.

2xYT medium:
16 g tryptone
10 g yeast extract
5 g sodium chloride (NaCl)
add distilled water to 1000 ml, autoclave at 121 °C for 15 min

Antibiotics: *B. subtilis* chloramphenicol (5 µg/ml)
E. coli ampicillin (50 µg/ml)

7. Transformation of *Bacillus subtilis*

Bacillus subtilis can be transformed via natural competence or by electroporation.

7.1. Protocol A - Natural Competence

The following protocol is adopted from Klein *et al.* (1992). Immediate usage of freshly prepared competent cells usually results in higher transformation rates.

Preparation of competent *Bacillus subtilis* cells

- Prepare an overnight culture of the appropriate recipient cells in 5 ml HS medium at 37 °C under vigorous shaking. For inoculation we recommend using one single colony grown on an LB agar plate.
- Measure the OD600 (optical density at 600 nm) of the overnight culture and inoculate 50 ml HS medium to an OD600 of 0.05; incubate under vigorous shaking at 37 °C
- Record the growth curve
- Immediately at transition of exponential to stationary growth phase start taking samples of 10 ml, each 15 min
- Add 1 ml of sterile glycerol (87%), mix and leave for 15 min on ice
- Fractionate into 1 ml aliquots, freeze in liquid nitrogen and store at -80 °C
- Check one aliquot from each time point with a reference plasmid DNA (see below) to identify the time point(s) yielding high level competent cells; discard the non- or low competent aliquots

Transformation of competent *Bacillus subtilis* cells

- Thaw one aliquot at 37 °C
- Use these cells to inoculate 20 ml LS medium
- Shake cells slowly in a 30 °C water bath to obtain maximal competence (about 2 h)
- Take 1 ml aliquots into a glass tube or 2 ml plastic reaction tube, add 10 µl of 0.1 M EGTA, and incubate for 5 min at room temperature
- Add 1 µg plasmid or chromosomal DNA and incubate for 2 h at 37 °C while well shaking (well mixing is important when using plastic reaction tubes)
- If glass tubes were used, transfer cell suspension into an plastic reaction tube
- Centrifuge, discard supernatant carefully and resuspend the cells into the final supernatant remaining on the pellet
- Plate on selective 2xYT medium containing 5µg/ml chloramphenicol
- Incubate at 37 °C overnight

7.2. Protocol B - Electroporation

Electroporation of *B. subtilis* (modified from Zhang *et al.*, 2011)

- Culture *B. subtilis* in 2xYT medium overnight
- Dilute 100-fold with 2xYT medium
- Grow culture to an OD600 of 0.2
- Then supplement culture with 1% DL-threonine, 2% glycine, 0.1% tryptophan and 0.03% Tween 80
- Grow while shaking for 1 h
- Cool on ice for 20 min
- Spin at 5000 x g for 10 min at 4 °C
- Wash twice with electroporation buffer
- Resuspend in electroporation buffer at 1/100 of the original culture volume
- Add 100 µl cells to an ice-cold 2 mm cuvette
- Add 2 µl DNA (25 ng/µl)
- Shock by a single 12.5 kV/cm pulse (Gene Pulser; Bio-Rad), resistance 200 Ω, capacitance 25 µF
- Immediately add 1 ml 2xYT broth containing 0.5 M sorbitol and 0.38 M mannitol
- Incubate at 37 °C for 3 h
- Spread on selective 2xYT plates

7.3 Media and Solutions

10x S-base (Spizizen's salt):
2 g $(NH_4)_2SO_4$
14 g K_2HPO_4
6 g KH_2PO_4
1 g sodium citrate
add distilled water to 100 ml and autoclave
add 0.1 ml 1M $MgSO_4$ after autoclaving

HS medium:
66.5 ml distilled water
10 ml 10x S-base
2.5 ml 20% (w/v) glucose
5 ml 0.1% (w/v) L-tryptophan
1 ml 2% (w/v) casamino acids
5 ml 10% (w/v) yeast extract (Difco)
10 ml 8% (w/v) arginine, 0.4% histidine
autoclave all components separately
tryptophan solution: sterile filtration

LS medium:
80 ml distilled water
10 ml 10x S-base
2.5 ml 20% (w/v) glucose
0.5 ml 0.1% (w/v) L-tryptophan
0.5 ml 2% (w/v) casamino acids
5 ml 2% (w/v) yeast extract (Difco)
0.25 ml 1 M $MgCl_2$
0.05 ml 1 M $CaCl_2$
autoclave all components separately
tryptophan solution: sterile filtration

0.1 M EGTA:	dissolve 3.8 g EGTA in 50 ml distilled water adjust the pH to 7.2 using 10 N NaOH add distilled water to 100 ml autoclave
2xYT medium:	16 g tryptone 10 g yeast extract 5 g sodium chloride (NaCl)
Electroporation buffer	0.5 M trehalose 0.5 M sorbitol 0.5 M mannitol 0.5 mM $MgCl_2$ 0.5 mM K_2HPO_4 0.5 mM KH_2PO_4 pH 7.4 filter-sterilize and store frozen

8. Induction with IPTG and Sample Analysis

- Grow appropriate *B. subtilis* strain overnight in fresh 2xYT medium
- Inoculate into fresh 2xYT medium to an OD_{600} of 0.15
- When culture reaches OD_{600} 0.7 – 0.8, split into 2 portions and induce with 1 mM IPTG to one portion (t = 0)
- Collect samples at different time points for analysis (t = 1, 2, 3,…)

8.1. Preparation of soluble and insoluble cell extracts from *B. subtilis*

- Harvest cells by centrifugation (10 min, 6,000 x g, 4 °C)
- Wash and resuspend in 50 mM sodium phosphate buffer (pH 7.0) at an OD_{600} of 10
- Disrupt cells by ultrasonication (12 W, 6 x 15 pulses with 15 sec intervals) in 1.5 ml Eppendorf tubes containing 1 ml of cell suspension, supplemented with lysozyme (250 µg/ml), on ice

- *Alternatively, cells can be disrupted by beat beating:*
  Disrupt three times with glass beads (0.1 mm in diameter) (1 g/ml of cell suspension) in an orbital mixer at 180 V, with the mix kept on ice for 3 min between each disruption

- Take 100 µl of the preparation as first total protein sample (T1)
- Remove remained cells by centrifugation at 4,300 x g, 10 min, 4 °C
- Take 100 µl of the supernatant for the second total protein sample (T2)
- Spin at 8.200 x g (10 min, 4 °C) to separate into insoluble (I) and soluble (S) protein fractions.
- Per sample use the amount of protein corresponding to 0.025 of OD_{600} for separation by SDS-PAGE
- Analyze samples by immunoblotting with specific antiserum

8.2. Precipitation of proteins from culture supernatant

- Collect proteins from cultured supernatant by TCA method
- Mix 1 volume of 40 % TCA with 3 volumes of culture supernatant
- Incubate on ice for 10 min
- Centrifuge at 12,000 x g at 4 °C for 10 min (until the supernatant is clear)
- Wash pellet twice with ice-cold acetone and dry at room temperature
- Dissolve pellet in 2x Laemmli buffer and denature at 95°C for 10 min for SDS-PAGE
  2x Laemmli: 125 mM Tris/HCl (pH 6.8), 4 % SDS, 20 % glycerine,
  10 % mercaptoethanol, 0.04 % bromphenol blue

9. References

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10. Order Information, Shipping and Storage

Order#	Product	Quantity
PBS013	pHT253 vector, with P*grac*100 and 8xHis tag at N-terminus	10 µg
PBS014	pHT254 vector, with P*grac*100 and 8xHis tag at C-terminus	10 µg
PBS015	pHT255 vector, with P*grac*100 and Strep tag at C-terminus	10 µg
PBS018	pHT1469 vector for secretory protein production, with P*grac*100, *amyQ** signal sequence and 8xHis tag at C-terminus	10 µg
Shipped at RT. Lyophilized plasmid DNA, can be stored at 4 °C. Once the DNA has been dissolved in sterile water or buffer we recommend storage at -20 °C.		
PBS020	*Bacillus subtilis* strain 1012wt	1 ml
PBS021	*Bacillus subtilis* strain 168 Marburg	1 ml
PBS022	*Bacillus subtilis* strain WB800N (for secretion vectors)	1 ml
PBS026	*Bacillus subtilis* strain AS1	1 ml
Shipped on dry ice; store at -80 °C		

11. Contact and Support

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These vector systems were initially constructed in the laboratory of Wolfgang Schumann at the Institute of Genetics, University of Bayreuth, Germany, Germany, and continue to develop in the laboratory of Hoang Duc Nguyen at the Center for Bioscience and Biotechnology, University of Science, Vietnam National University, Ho Chi Minh City.

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